

## APPENDIX 13.2 – A: Bioaerosol Sampling Methods and Limitations

Note: We update the information resource links at least semi-annually, but please let us know if you find a link that has changed by sending an email to [IH\\_Information\\_Systems@nehc.mar.med.navy.mil](mailto:IH_Information_Systems@nehc.mar.med.navy.mil).

VIABLE AND NON-VIABLE BIOAEROSOL SAMPLING METHODS				
Collection Method	Sample Equipment	Analysis Method(s)	Sampling Notes	Information Resource
Impaction into liquid	<b>AGI-30</b>  (all-glass impinger)	Microscope Culture Bioassay Chemical assay	12.5 LPM $d_{50} = 0.31\mu\text{m}$ ; particle collection efficiency $\approx 70\%$ @ $0.5\mu\text{m}$ ; $\approx 90\%$ @ $1\mu\text{m}$ Sample collection up to 2 hours Liquid can evaporate or spill. Use sterile oil (mineral or glycerol) to prevent evaporation. CANNOT use glycerol for bacterial recovery. Spores can re-aerosolize during collection	<a href="http://www.aceglass.com">www.aceglass.com</a> See Impinger #7540
Impaction into liquid	<b>BioSampler®</b>  (aka SAC = swirling aerosol collector)	Culture (hardy) Microscope Chemical assay Immunoassay Bioassay	12.5 LPM Long term collection (up to 8 hrs) with mineral oil Reduced particle bounce Minimal re-aerosolization Liquid can evaporate or spill. Use mineral oil or glycerol to eliminate evaporation (glycerol for PCR & microscope analysis; sterile mineral oil for fungus & bacterial culture). Particle collection efficiency $\approx 85\%$ @ $0.5\mu\text{m}$ ; $\approx 100\%$ @ $1\mu\text{m}$	<a href="http://www.skincinc.com/prod/Bio_sampler.asp">http://www.skincinc.com/prod/Bio_sampler.asp</a>
Impaction onto stainless or filter	<b>Andersen Non-Viable 8-stage</b>  (cascade impactor)	Microscope	Aerosol can impact directly onto collection surface or onto filtration media Allows size selection. Cut points at standard collection of 28.3 LPM = 0.4, 0.7, 1.1, 2.1, 3.3, 4.7, 5.8, & 9.0 $\mu\text{m}$ Additional special stages allow 60 & 90 LPM collection for sub-micron particle collection.	<a href="http://www.anderseninstruments.com/eight_stage.htm">http://www.anderseninstruments.com/eight_stage.htm</a>
Impaction onto stainless or filter	<b>Andersen Personal Cascade Impactor</b>  (personal BZ)		Collect at 2 LPM 8-stage impactor  $d_{50}$ stage 1 = 21; stage 2 = 14.8; stage 6 = 1.55; stage 8 = 0.52 $\mu\text{m}$	<a href="http://www.anderseninstruments.com/marple_290_impactor.htm">http://www.anderseninstruments.com/marple_290_impactor.htm</a>
Impaction onto agar	<b>Aerotech 6™</b>	Culture	See Andersen N-6	<a href="http://www.aerotechlabs.com/Aero/">http://www.aerotechlabs.com/Aero/</a>

## VIALE AND NON-VIALE BIOAEROSOL SAMPLING METHODS

Collection Method	Sample Equipment	Analysis Method(s)	Sampling Notes	Information Resource
Impaction onto agar	<b>Andersen N-6</b>	Culture	<p>Precise speciation; differentiates between <i>Aspergillus</i> &amp; <i>Penicillium</i></p> <p>Quantification (CFU/m<sup>3</sup>)</p> <p>Collected using high volume pump and agar plates</p> <p>Standard collection is 28.3 LPM for 5 minutes (snapshot sample)</p> <p>Allows size selective sampling based on stages used</p> <p>d<sub>50</sub> stage 1 = 6.61µm; d<sub>50</sub> stage 6 = 0.57µm</p> <p>Reports only viable spores (underestimates total spores)</p> <p>5-20 day incubation</p> <p>Requires sterile collection</p> <p>Routine collection misses species with special growth requirements (e.g., <i>Stachybotrys</i>)</p> <p>Requires positive hole correction</p> <p>Requires disinfection between samples to prevent cross-contamination</p>	<a href="http://www.anderseninstruments.com/viable_samplers.htm">http://www.anderseninstruments.com/viable_samplers.htm</a>
Impaction onto agar	<b>Biocassette</b> (personal BZ)		Single use sampler (disposable)	<a href="http://www.emlab.com/media/resources/biocassette.pdf">www.emlab.com/media/resources/biocassette.pdf</a>
Impaction onto agar	<b>BioStage®-1</b>  Standard BioStage®  Micromedia BioStage®	Culture	<p>Replica of the Andersen N-6 bioaerosol sampler. Catalog 225-9535.</p> <p>Single stage; 28.3 LPM; 400 holes; 90-100 mm plates. Catalog 225-9611.</p> <p>Single stage; 14. 15 LPM; 200 holes; 60 mm plates. Catalog 225-9610.</p>	<a href="http://www.skinc.com">www.skinc.com</a>  <a href="http://www.skinc.com/prod/225-9611.asp">http://www.skinc.com/prod/225-9611.asp</a>
Impaction onto agar	<b>Burkard Portable Air Sampler</b>	Culture	Sampling time 1-9 minutes (or continuous) at 10 LPM 90 mm agar plates	<a href="http://www.burkard.co.uk/portsamp.htm">http://www.burkard.co.uk/portsamp.htm</a>
Impaction onto agar	<b>M Air T Tester</b> (sieve impactor)		<p>Collect at 140-180 LPM; maximum sample volume 1000 L</p> <p>d<sub>50</sub> = 0.3.5 µm</p> <p>Usually used for clean rooms</p> <p>Sieve collection allows uniform distribution onto agar</p>	<a href="http://www.millipore.com/publications.nsf/docs/PF10176EN00">http://www.millipore.com/publications.nsf/docs/PF10176EN00</a>
Impaction onto agar	<b>Mattson-Garvin M/G Air Sampler 220</b>  (slit to agar)	Culture	<p>Agar plate on rotating surface, sampled at 28.3 LPM</p> <p>d<sub>50</sub> = 0.5 µm</p> <p>Drive motors can be changed to set 1 revolution of the plate at 5, 15, 30, or 60 minutes.</p>	<a href="http://www.mattson-garvin.com/models.htm">http://www.mattson-garvin.com/models.htm</a> (Mattson-Garvin, Barramundi Corporation)

## VIABLE AND NON-VIABLE BIOAEROSOL SAMPLING METHODS

Collection Method	Sample Equipment	Analysis Method(s)	Sampling Notes	Information Resource
Impaction onto agar	<b>MicroBio MB2</b>	Culture	Portable Standard sample rate 100 LPM; adjustable sample time Total air volume 25-1000 L Laminar air flow through 1 mm holes onto 55 mm contact agar plate	<a href="http://www.aicompanies.com/SBI_products/airsample/MB2/mb2.htm">http://www.aicompanies.com/SBI_products/airsample/MB2/mb2.htm</a>
Impaction onto agar	<b>RCS</b>  (Reuter Centrifugal Sampler)  (collection onto agar strips)	Culture	<u>Standard RCS</u> – Sampling times 0.5, 1, 2, 4, 8 minutes  <u>RCS Plus</u> – Typical collection at 50 LPM for 20 minutes; maximum sample volume 2000L. $d_{50} \approx 2-5 \mu\text{m}$  <u>RCS High Flow</u> – Typical collection 100 LPM for 10 minutes; maximum sample volume 1000L. $d_{50} \approx 2-5 \mu\text{m}$  <u>RCS Isolator</u> – Flow rate 100 LPM; maximum volume 1000 L	<a href="http://www.biotestusa.com/sampling.html">http://www.biotestusa.com/sampling.html</a>
Impaction onto agar	<b>SAS</b>  (Surface Air System)	Culture	Collect at 90-180 LPM, using 55, 84, or 90 mm agar plates $d_{50} = 0.67\mu\text{m}$ $d_{50} = 1.45\mu\text{m}$ Requires hole correction SAS Super 100 (100 LPM) can be programmed for collection time and volume	Bioscience International, <a href="http://www.biosci-intl.com/products/sas100.htm">http://www.biosci-intl.com/products/sas100.htm</a>
Impaction onto agar	<b>SPS 3000</b>	Culture	Cascade impaction using 2, 3, or 6 stages. 6 stage hole diameters: stage 1 – 1.20 mm; stage 2 – 0.90 mm; stage 3 – 0.70 mm; stage 4 – 0.50 mm; stage 5 – 0.35 mm; stage 6 – 0.25 mm	<a href="http://www.lanzoni.it/">http://www.lanzoni.it/</a> Enter site – Choose ‘Enter’ over British flag – Choose Products & Services
Impaction onto agar	<b>STA-203/ 204</b>  (slit to agar collection onto rotating agar surface)	Culture	Sample exposure time (i.e., time for 1 revolution of agar plate) can be adjusted from 2-100 minutes  <u>STA 203</u> - Flow rate 15-50 LPM (30-5000 L total sample volume)  <u>STA 204</u> - Flow rate 15-30 LPM (30-3000 L total sample volume) Includes built-in vacuum pump	<a href="http://www.nbsc.com">www.nbsc.com</a> (New Brunswick Scientific – Choose Biological Air Samplers from the Products list)
Impaction onto agar	<b>Zefon A6</b>	Culture	$d_{50} = 0.65 \mu\text{m}$ (1-stage only available)	<a href="http://www.zefon.com">www.zefon.com</a>

## VIALE AND NON-VIALE BIOAEROSOL SAMPLING METHODS

Collection Method	Sample Equipment	Analysis Method(s)	Sampling Notes	Information Resource
Impaction onto tape or slide	<b>Air-O-Cell™ cassette</b>  (sticky sampling media)	Microscope	Quick, easy to use Collect viable & non-viable particles using high volume pump & Air-O-Cell cassette (37 mm) Standard sampling is 15 LPM for 10 minutes; flow rate range 5-30 LPM $d_{50} = 2.6 \mu\text{m}$ @ 15 LPM Particle collection efficiency @ 15 LPM $\approx 0$ @ $2\mu\text{m}$ ; 85% @ $3\mu\text{m}$ ; 95% @ $15\text{-}80\mu\text{m}$ Can get identification & quantification (spores/ $\text{m}^3$ & speciation) Cassettes average \$7-\$10 each; long shelf life Specially adapted pump available from Zefon (MiniPump) & from SKC (QuickTake 15)	<a href="http://www.zefon.com">www.zefon.com</a>  <a href="http://www.skinc.com/prod/225-9501.asp">http://www.skinc.com/prod/225-9501.asp</a>
Impaction onto tape/ slide	<b>Burkard Continuous Recording Personal Air Sampler</b>	Microscope	Personal sampling continuously over 24 hour period Particle collection efficiency $\approx 90\%$ for particles $\geq 5\mu\text{m}$ Preset selection for 2 mm/hr for 24 hr; 4 mm/hr for 12 hrs; or 8 mm/hr for 6 hr Allows timed sample collection over 24-hr intervals	<a href="http://www.burkard.co.uk/containers.htm">http://www.burkard.co.uk/containers.htm</a>
Impaction onto tape/ slide	<b>Burkard Spore Trap</b>  (personal sampler on moving slide)	Microscope	$d_{50} = 2.52 \mu\text{m}$ ; very poor collection efficiency of particles $< 2 \mu\text{m}$ Poor particle deposition uniformity Collect at 10 LPM for 1-9 minutes	<a href="http://www.burkard.co.uk/perssamps.htm">http://www.burkard.co.uk/perssamps.htm</a>
Impaction onto tape/ slide	<b>Burkard Spore Trap – 7 day Recording Sampler</b>  (rotating drum)	Microscope	Tape/slide on rotating drum, allowing 1-7 days continuous recording 10 LPM - drum rotates 1 revolution/7 days @ 2mm/hour. Allows timed sample collection to follow fluctuations over 24-hr interval. $d_{50} = 5.2 \mu\text{m}$ ; Can improve collection efficiency of particles $< 10 \mu\text{m}$ by changing orifice. Rugged equipment. Usually used for outdoor sampling, especially air quality testing, e.g., pollen counts.	<a href="http://www.burkard.co.uk/7days.htm">http://www.burkard.co.uk/7days.htm</a>
Impaction onto tape/ slide	<b>Cyclex™</b>  (circular bioaerosol impact sampler)	Microscope	$d_{50} = 1.8 \mu\text{m}$ Collect at 20 LPM for up to 10 minutes Collects non-viable samples using aluminum $360^\circ$ impaction chamber.. Can also be used for collecting carpet and cavity samples with accessories.	<a href="http://www.emssales.net">http://www.emssales.net</a>

## VIALE AND NON-VIALE BIOAEROSOL SAMPLING METHODS

Collection Method	Sample Equipment	Analysis Method(s)	Sampling Notes	Information Resource
Impaction onto tape/ slide	<b>Cyclex-D™ cassette</b>	Microscope	Bioaerosol sampling for fungal spores, pollen, fibers, dander, and others. Used with a high volume pump; 20 LPM for 1-15 minutes Impacts onto transparent adhesive slide in the center of the cassette. Particle collection efficiency $\approx 65\%$ @ $1\mu\text{m}$ ; $82\%$ @ $2\mu\text{m}$ ; $98\%$ @ $3\mu\text{m}$ $d_{50} \approx 1\mu\text{m}$	<a href="http://www.emssales.net">http://www.emssales.net</a>
Impaction onto tape/ slide	<b>Micro5 Microcell</b> (personal sampler)	Microscope	Personal and area sampling; 5 LPM for 1-15 minutes $d_{50} \approx 1\mu\text{m}$ Particle collection efficiency $\approx 89\%$ @ $1\mu\text{m}$ ;	<a href="http://www.emssales.net">http://www.emssales.net</a>
Impaction onto tape/ slide	<b>MK-3</b> (moving slide)	Microscope	Sample collected on greased slide at 15 LPM $d_{50} = 2.0\mu\text{m}$	Purchase <a href="http://www.emssales.net/productlist.aspx?CategoryID=2&amp;SubID=7">http://www.emssales.net/productlist.aspx?CategoryID=2&amp;SubID=7</a> Information <a href="http://www.gapenviromic.com/images/MK_3_Air_Sampler_Operation.pdf">http://www.gapenviromic.com/images/MK_3_Air_Sampler_Operation.pdf</a>
Impaction onto tape/ slide	<b>Rotorod</b> (tape on rotating rod)	Microscope	Typically used for air quality monitoring (pollen counts)	<a href="http://www.multidata.com/ProductCatalog.html">http://www.multidata.com/ProductCatalog.html</a>
Impaction onto tape/ slide	<b>VPPS 1000</b> (volumetric particle sampler using rotating drum)	Microscope	10 LPM for 24-hour sampling period (slide moves at 2 mm/hr) Can be set for 1-7 days continuous recording	<a href="http://www.lanzoni.it">www.lanzoni.it</a> Enter site – Choose ‘Enter’ over British flag – Choose Products & Services
Impaction onto tape or slide	<b>WallChek™</b> (wall cavity sampling)	Microscope	Provides essentially non-destructive sampling in wall cavities (requires a 3/8" hole). A special adaptation using an Air-O-Cell cassette.	<a href="http://www.aerotechlabs.com/Aero">www.aerotechlabs.com/Aero</a> <a href="http://www.skincinc.info/instructions/37504.pdf">http://www.skincinc.info/instructions/37504.pdf</a>

## VIALE AND NON-VIALE BIOAEROSOL SAMPLING METHODS

Collection Method	Sample Equipment	Analysis Method(s)	Sampling Notes	Information Resource
Filtration	<b>Button™ Aerosol Sampler</b>  (inhalable mass)	Culture Microscope	25 mm filter; 4 LPM sample collection; Collects inhalable fraction Reduces electrostatic effects Good filter collection uniformity Particle collection efficiency ≈100% for 0.5 – 5 µm particles when sampling long term (some sensitive species may dessicate). Good for collecting bacteria if using gelatin filter	<a href="http://www.skincinc.com/prod/225-360.asp">http://www.skincinc.com/prod/225-360.asp</a>
Filtration	<b>Cassette</b> 25 mm 37 mm 45 mm	Microscope Culture (hardy species) Bioassay Chemical assay Immunoassay	Usually collected at 1-5 LPM Particle collection efficiency good at 0.5µm and above Sterile filters required for culturing Filters with grids can be purchased for counting Electrostatic charges may be collection problem Filtration methods tend to concentrate particles at center of filter Good collection method for long-term sampling of hardy spores	<a href="http://www.zefon.com">www.zefon.com</a>  <a href="http://www.skincinc.com/prod/gel_filters.asp">http://www.skincinc.com/prod/gel_filters.asp</a>
Filtration	<b>IOM Sampler</b>	Culture Microscope	Use sterilized 25 mm polycarbonate filters to collect personal bioaerosol samples. Use multidust foam discs to size select into inhalable, thoracic, and respirable fractions. Flow rate 2 LPM	<a href="http://www.skincinc.com/prod/225-70A.asp">http://www.skincinc.com/prod/225-70A.asp</a>
Filtration	<b>Laro-100</b>	Culture Microscope	Uses 0.8 µm MCEF filters. Claims to capture 100% of particles from 1-10 µm. 4-5 LPM for 3-30 minutes. Can be used for 8-hr personal sample collection at 1-2 LPM flow rate.	<a href="http://www.laro100.com/">http://www.laro100.com/</a>  <a href="http://www.emsl.com/laro_sampling_guide.html">http://www.emsl.com/laro_sampling_guide.html</a>

## SURFACE AND BULK SAMPLING METHODS

Collection Method	Sample Equipment	Analysis Method(s)	Sampling Notes	Information Resource
Bulk	Bulk material or dust	Cultured or direct microscope exam  CFU/g	Carpet or dust sampling 25 mm (with conductive cowl) or 37 mm cassettes with beveled inlet port	Carpet sampling pump kit - <a href="http://www.skcinc.com/prod/225-9541.asp">http://www.skcinc.com/prod/225-9541.asp</a>  MicroVac - <a href="http://www.zefon.com">www.zefon.com</a>
Swab	Bulk material or dust with sterile swab (Q-tip or cotton ball will work)	Direct microscope or culture  CFU/m <sup>2</sup>	Quick Inexpensive Measured template required for quantification Surfaces may not adequately characterize extent or type of contamination No correclation to airborne results  Swab & template kits available	<a href="http://www.zefon.com">www.zefon.com</a>  <a href="http://www.skcinc.com/prod/sterilswab.asp">http://www.skcinc.com/prod/sterilswab.asp</a>
Tape	Clear sticky tape placed directly over the surface to be sampled.  Place tape onto slide for transport	Microscope	Quick Inexpensive Kits available, e.g., BioTape™ (flexible plastic slide with adhesive center from Zefon),	Zefon <a href="http://www.zefon.com">http://www.zefon.com</a>

## OTHER MICROBIOLOGICAL COLLECTION METHODS

Resource Information Available from Laboratories Offering These Tests (See [Laboratory Services Chart, Appendix C](#))

Analyte	Sample Information	Analysis Method(s)	Sampling Notes
<b>Allergens</b>	Impinger methods Filter collection followed by Wash Settled dust	Immunoassay (IA)  (antigen-antibody specific)	Analyzes viable & non-viable for available antigenic species. Quick analysis. Can be fluorescent IA, enzyme IA [ELISA], or radio IA [RAST] Special equipment & expertise needed. Cross-reactivity can occur. Typical sample allergens = dog, cat cockroach, dust mites
<b>β glucan</b> (1→3)-β-D glucan  (fungal cell wall component)	Filtration: 0.8μm cellulose acetate/ nitrate filter or 1 μm polycarbonate	Chemical assay or Bioassay (LAL)	β glucan is a cell wall component of all fungi; sampling can be used as biomarker for the presence of any fungal contamination Detects dead & living spores Does not identify which fungal species is present Quick analysis time
<b>Endotoxins</b>  (Gram negative bacteria)	Filtration: 37 mm cassette with polycarbonate filter  Can use bulk sample	Bioassay (LAL)  Chemical assay via GC-MS or HPLC  Kinetic chromogenic assay	Cassettes/filters/vials must be sterile and endotoxin-free Identifies toxins produced by gram negative bacteria Can analyze air, dust, or bulk material samples (including water samples). May be useful as a post-remediation sample after a sewage spill or backup. Have shown correlation between occupant symptoms & endotoxin concentrations
<b>EPS</b> (bacteria & fungi)		Immunoassay	EPS = Extracellular polysaccharides
<b>Ergosterol</b>  (fungal cell wall component)	Filtration  0.45μm polycarbonate filter (closed face)  1-4 LPM	Chemical assay (HPLC, MS or GC-MS)	Ergosterol is the main membrane sterol in most fungi; sampling is used as biomarker for the presence of fungal contamination Detects dead & living spores Long-term stability before analysis Does not identify which fungal species is present Reported in μg ergosterol
<b>Fatty Acids</b>  (bacteria)		Chemical assay	Fatty acids are cell wall components of bacteria. Analysis uses gas chromatography to determine the fatty acid profile, then compares results to a reference database using statistical pattern recognition software.
<b>MVOCs</b>  (Microbial Volatile Organic Compounds)	Low flow pump with sorbent tube (e.g., Anasorb 747 carbon)  0.2 LPM for 240 minutes	Chemical assay (GC-MS)	Mold must be actively growing for organic compound production. Samples must be frozen after collection. Samples can be collected using summa canisters. Little consensus on which MVOCs are “important” or medically significant.
<b>Mycotoxins</b>  (Fungi)	Filtration: 37 mm cassette With 1 μm PTFE (#003-2028-00)	Chemical assay (HPLC; GC/MS; LC/MS)	Sensitive analytical methods not available for most indoor air mycotoxins. Agriculturally important toxins methods widely available. Identifies secondary metabolites of fungi, produced only when the fungus is stressed.



## OTHER MICROBIOLOGICAL COLLECTION METHODS

Resource Information Available from Laboratories Offering These Tests (See [Laboratory Services Chart, Appendix C](#))

Analyte	Sample Information	Analysis Method(s)	Sampling Notes
<b>PCR Genetic Identification</b>	Filter: 37 mm cassettes with 1 µm PTFE filter	Bioassay, i.e., PCR (polymerase chain reaction)	Genetic identification using genus or species-specific DNA primers Analysis within 1 day Expensive; requires expertise & special equipment DNA primers limited (mostly <i>Aspergillus</i> , <i>Penicillium</i> , <i>Stachybotrys</i> )

### Abbreviations Key:

β	beta	GC	gas chromatography	LAL	<i>Limulus</i> amoebocyte lysate assay
BZ	breathing zone	MS	mass spectrometry	PTFE	polytetrafluoroethylene
m <sup>3</sup>	cubic meters	HPLC	high performance liquid chromatography	PCR	polymerase chain reaction
≈	approximately	mm	millimeters	LC	liquid chromatography
µm	micrometers				
LPM	liters per minute				
MVOC	microbial volatile organic compounds				
d <sub>50</sub>	cut-off diameter or cut size; the aerodynamic particle size at which 50% of the particles will be captured by the collection method of interest.				

### Analysis Methods:

Culture (viable) – collect or transfer sample onto nutrient agar to grow colonies  
 Microscope (non-viable) – examine under microscope for counting and/or morphological identification  
 Bioassay – analytical method in which the result is an observable effect on or in a biological system/ organism. Includes genetic identification using DNA primers (i.e., polymerase chain reaction (PCR)) and LAL assay for endotoxins.  
 Chemical Assay – analysis of chemical compounds produced by or contained in the microbe, i.e., MVOCs, mycotoxins  
 Immunoassay – analysis based on the specificity of an antigen-antibody reaction (e.g., cat, dog, cockroach, & dust mite allergen tests; ELISA)

### Culture:

#### Advantages

Species identification  
 Large reference database  
 No special equipment needed

#### Disadvantages

Slow results (7-10 days incubation)  
 Media is selective  
 Underestimates total count  
 Detects only living organisms that are able to live & compete on selected nutrient media & conditions  
 Species cultured from a site may not be the most prevalent or important

### Microscope:

Analyzes total count  
 Fast  
 Affordable

Limited identification ability; cannot distinguish some species  
 Background debris can interfere with identification  
 Special expertise needed for confident identification

### Biological & Chemical Assay

Fast  
 Analyzes viable & non-viable  
 Can screen for specific genus/species

Special equipment & expertise needed  
 Limited reference data except for endotoxins